
Biomimetic cyclodextrin nanotubes for Ion-channel Applications: Design, synthesis and ionic conduction

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Abstract

For few years, we observe the development of biomimetic channels for DNA sequencing such as carbon nanotubes, inserted into bilayers in despite of their hydrophobic properties^{1,2}, or aerolysin nanopore for the discrimination of polymers³ or oligonucleotides⁴ of different lengths. Recently, biomimetic synthetic channels were developed based on DNA duplex⁵ or more complex DNA-based origamis^{6,7,8,9} to design new tools for DNA sequencing or barcode based biomarkers¹⁰. These new channels will own a high confinement due to their thin internal diameter without using a cyclodextrin based adaptor^{11,12,13}. Based on our knowledge of modified cyclodextrin channels¹⁴, we synthesize biomimetic cyclodextrin nanotubes while keeping a cylindrical confined geometry¹⁵. As synthesis of these nanotubes is precisely controlled, we modify their diameter by using α -, β - or γ -cyclodextrins, their lengths with a monodisperse distribution according to TEM imaging and their functionalization to enhance their life time. We show that the conductance of these sub-nanometer nanotubes is governed by the same ionic transport as in gramicidin channels. These synthetic channels are not cytotoxic, leading to biomedical applications.

¹ Geng, J. *et al. Nature* **2014**, *514*, 612–615.

² Liu, L.; Yang, C.; Zhao, K.; Li, J.; Wu, H.-C. *Nat. Commun.* **2013**, *4*.

³ Baaken, G.; Halimeh, I.; Bacri, L.; Pelta, J.; Oukhaled, A.; Behrends, J. C. *ACS Nano* **2015**, *9*, 6443–6449.

⁴ Cao, C.; Ying, Y.-L.; Hu, Z.-L.; Liao, D.-F.; Tian, H.; Long, Y.-T. *Nat. Nanotechnol.* **2016**, *11*, 713–718.

⁵ Goepfrich, K. *et al. Nano Lett.* **2016**, *16*, 4665–4669

⁶ Bell, N. A. W.; Engst, C. R.; Ablay, M.; Divitini, G.; Ducati, C.; Liedl, T.; Keyser, U. F. *Nano Lett.* **2012**, *12*, 512–517.

⁷ Langecker, M. *et al. Science* **2012**, *338*, 932–936

⁸ Burns, J. R.; Seifert, A.; Fertig, N.; Howorka, S. A *Nat. Nanotechnol.* **2016**, *11*, 152–156

⁹ Goepfrich, K. *et al. ACS Nano* **2016**, *10*, 8207–8214

¹⁰ Zhang, X.; Wang, Y.; Fricke, B. L.; Gu, L.-Q. *ACS Nano* **2014**, *8*, 3444–3450.

¹¹ Astier, Y.; Braha, O.; Bayley, H. *J. Am. Chem. Soc.* **2006**, *128*, 1705–1710.

¹² Deamer, D.; Akeson, M.; Branton, D. *Nat. Biotechnol.* **2016**, *34*, 518–524.

¹³ Fuller, C. W. *et al. Proc. Natl. Acad. Sci. U. S. A.* **2016**, *113*, 5233–5238.

¹⁴ Bacri, L.; Benkhaled, A.; Guegan, P.; Auvray, L. *Langmuir* **2005**, *21*, 5842–5846.

¹⁵ Mamad-Hemouch, H. *et al. Nano Lett.* **2015**, *15*, 7748–7754